

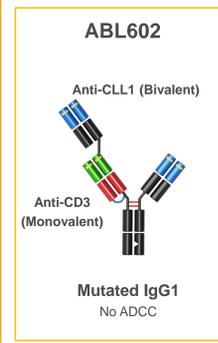
A novel asymmetrical anti-CLL-1×CD3 bispecific antibody, ABL602, induces potent CLL1-specific antitumor activity with minimized sensitization of pro-inflammatory cytokines

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INTRODUCTION

- Acute myeloid leukemia (AML) is a disease with high incidence of relapse that is originated and maintained from leukemia stem cells (LSCs).
- Expression of C-type lectin-like molecule-1 (CLL-1; also known as CLEC12A, c-type lectin domain family 12 member A) is mainly restricted to LSCs but absent in normal hematopoietic stem cells (HSCs).
- This unique CLL-1 expression pattern paves the way to develop therapies that potentially eliminate CLL-1-positive LSC while preserving CLL-1-negative HSC.

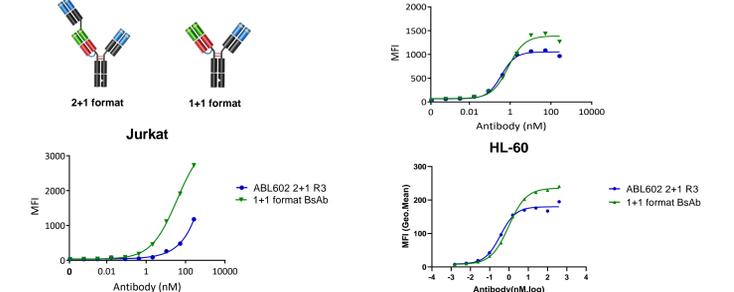
SUMMARY



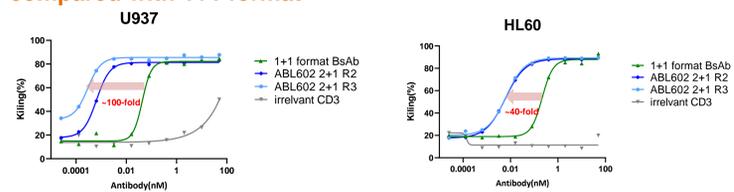
- ABL602 has bivalent binding arms to CLL-1 as a cancer target and a monovalent binding arm to CD3.
- ABL602 exhibited higher binding activity to CLL1-expressing AML cell lines and greater tumor-killing efficacy than 1+1 format BsAb and benchmark antibody MCLA-117.
- ABL602 induced potent cytotoxic activities on CLL1-expressing AML cell lines with concomitant T cell activation and cytokine/granzyme B release.
- ABL602 did not or minimally induced TNF- α and IL-6 in PBMC in the absence of AML cell lines, while MCLA-117 triggered high level of expression of those cytokines.
- In established orthotopic AML mouse model using HL-60 Luc, ABL602 demonstrated statistically significant anti-tumor activity in a dose-dependent manner.

RESULT

ABL602 2+1 shows distinct target binding properties compared with 1+1 format



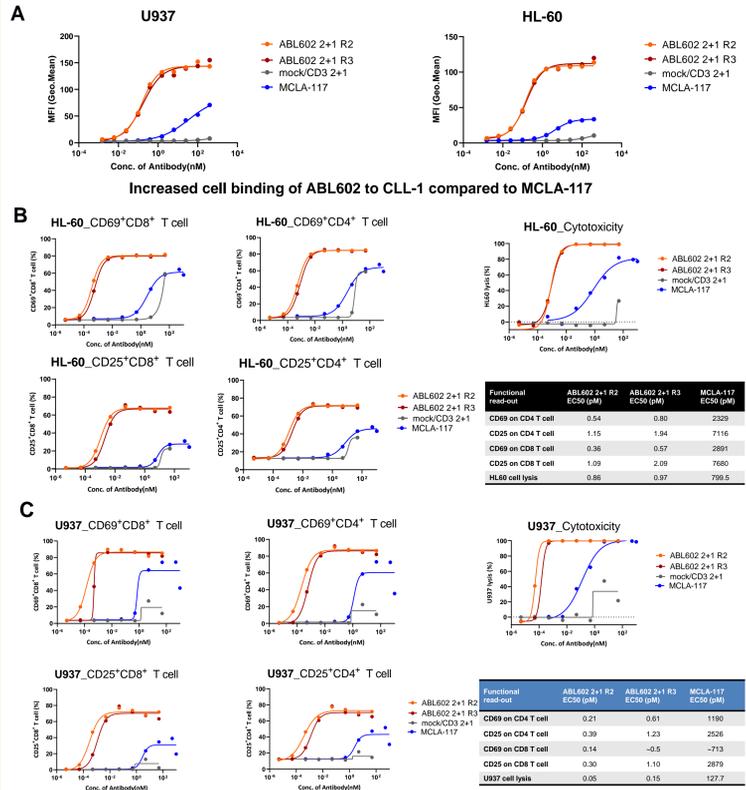
ABL602 2+1 induces potent cytotoxic activity on AML cell lines compared with 1+1 format



Activity of ABL602 was assessed with T cell-dependent cellular cytotoxicity (TDCC) assays. PBMC were incubated with target cells with ET ratio of 10:1 and ABL602 2+1 or 1+1 format. Cytotoxicity was assessed by flow cytometry after 48 h incubation at 37°C. Two T cell donors were tested per cell line, in 2 independent experiments. R represents G4S linker.

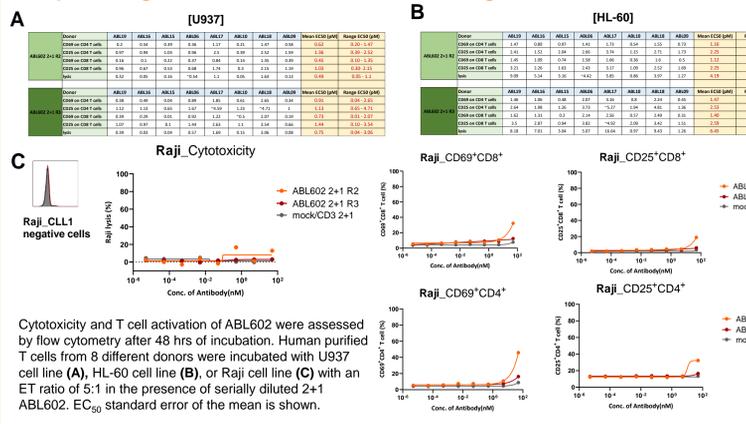
RESULT

ABL602 induces much stronger T cell activation and cytotoxicity on CLL1-positive tumor cells than MCLA-117



A.U937 and HL-60 cell lines were stained with various concentrations of ABL602, or MCLA 117 to characterize the surface-binding profiles of the bispecific antibody. HL-60 (B) or U937(C) suspension target cells (2 x 10⁴ cells) were seeded onto 96-well U bottom plates. Various concentrations of ABL602, mock/CD3, or MCLA-117 and human purified T cells were added to the plates at an effector:target ratio of 5:1. After 48hrs, T cell activation and the number of remaining CD3 positive target cells were quantified by flow cytometry. T cells were analyzed by flow cytometry for cell surface levels of CD25 and CD69 as a markers of T cell activation.

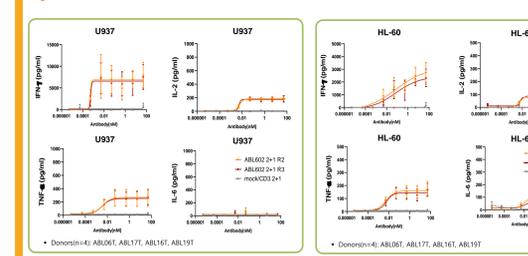
ABL602 induces potent T cell activation and cytotoxicity on CLL1-expressing cell lines but not on CLL1-negative cell lines



Cytotoxicity and T cell activation of ABL602 were assessed by flow cytometry after 48 hrs of incubation. Human purified T cells from 8 different donors were incubated with U937 cell line (A), HL-60 cell line (B), or Raji cell line (C) with an ET ratio of 5:1 in the presence of serially diluted 2+1 ABL602. EC₅₀ standard error of the mean is shown.

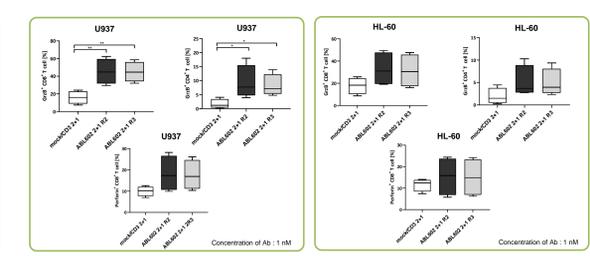
RESULT

ABL602 induces cytokine expression in the presence of U937 and HL-60 cell lines



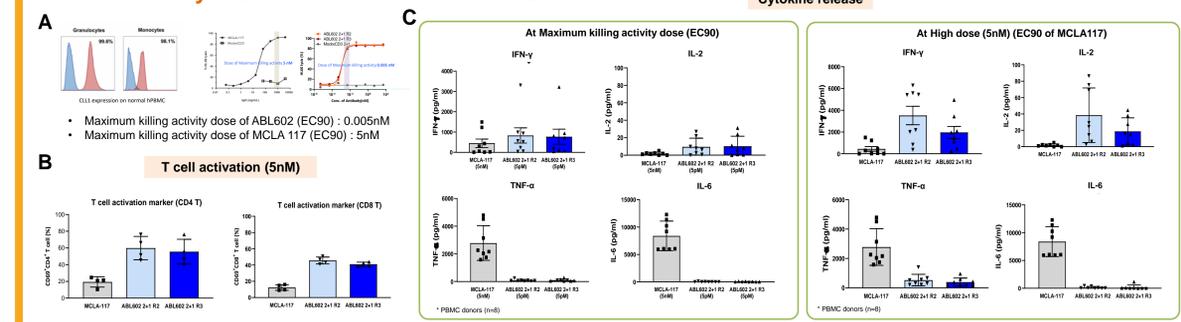
The purified T cells were treated with ABL602 2+1 either with U937 cell line or HL60 cell line for 48 h. The purified T cells were added at a 5:1 effector to target cell ratio. Supernatant was harvested to measure the human cytokine levels using ELISA. Cytokine levels were detectable for the four cytokines shown.

ABL602 induces Granzyme B and Perforin in the presence of U937 and HL-60 cell lines



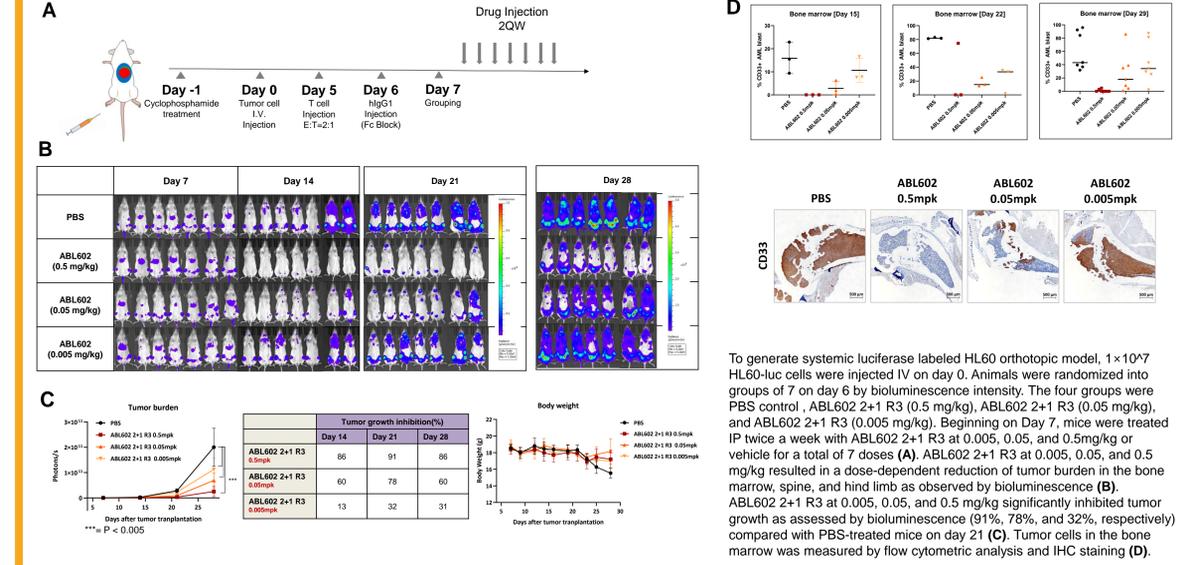
Human T cells are isolated from healthy PBMCs and then incubated with tumor cells and ABL602 2+1 or mock/CD3 2+1 for 24 h. After 5 hours of treatment with Golgi-plug/protein transport inhibitor, granzyme B and perforin protein were measured by flow cytometry. * : P<0.5, ** : P<0.05

ABL602 strongly induces T cell activation and cytokine expression of IFN-gamma and IL-2 but weakly CRS-related cytokines such as TNF-alpha and IL-6



Both granulocytes and monocytes from peripheral blood cells express CLL1 antigen (A, left). Dose curves show cytotoxicity of ABL602 or MCLA 117 in HL-60 cell line (A, right). Healthy donor-derived PBMCs were cultured with ABL602 2+1 or MCLA-117 antibody for 48 hours. T cells were analyzed by flow cytometry for cell surface levels of CD25 and CD69 as a markers of T cell activation (B). Supernatant was harvested to measure the human cytokine levels in these supernatants using ELISA. Cytokine levels were detectable for the four cytokines shown (C).

ABL602 inhibits tumor growth in a dose-dependent manner in HL-60 orthotopic model



To generate systemic luciferase labeled HL60 orthotopic model, 1x10⁷ HL60-luc cells were injected IV on day 0. Animals were randomized into groups of 7 on day 6 by bioluminescence intensity. The four groups were PBS control, ABL602 2+1 R3 (0.5 mg/kg), ABL602 2+1 R3 (0.05 mg/kg), and ABL602 2+1 R3 (0.005 mg/kg). Beginning on Day 7, mice were treated IP twice a week with ABL602 2+1 R3 at 0.005, 0.05, and 0.5mg/kg or vehicle for a total of 7 doses (A). ABL602 2+1 R3 at 0.005, 0.05, and 0.5 mg/kg resulted in a dose-dependent reduction of tumor burden in the bone marrow, spine, and hind limb as observed by bioluminescence (B). ABL602 2+1 R3 at 0.005, 0.05, and 0.5 mg/kg significantly inhibited tumor growth as assessed by bioluminescence (91%, 78%, and 32%, respectively) compared with PBS-treated mice on day 21 (C). Tumor cells in the bone marrow was measured by flow cytometric analysis and IHC staining (D).